# Establishing a Faunal Comparative Collection to Aid Cultural Resource Interpretation of Sites in Western Washington

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#### INTRODUCTION

Analysis of fauna in the archaeological record contributes to more in-depth understanding of the anthropological relationship people had with animals in the past. To increase access and understanding of fauna associated with cultural material, Antiquity Consulting is creating a basic faunal comparative collection from salvaged remains of animals common in western Washington. By paying close attention to the bone's surface, pathologies, and context, a more holistic approach can be made during archaeological surveys. The goal of the project is to create an ethical faunal comparative collection of common native and nonnative animals in western Washington to enrich analysis of cultural resources and provide local access to a collection focused on fauna.

## BACKGROUND

In the area now known as western Washington, relationships people had with animals are evident in associated with archaeological assemblages can paint a more holist picture of the people in the area. the archaeological record. Mammal, fish, birds, and shellfish hold key evidence of how humans Faunal remains have also been associated with European settlers from the early 19th century to the interacted with animals, in tandem with oral traditions and other cultural materials in context. In the present. Europeans brought livestock, pets, and invasive species with them as they traveled west, Pacific Northwest, many indigenous groups sustainably harvested and preserved salmonid for over attributing certain dates of introduction. The presence of some of these animals at archaeological sites 10,700 years as the Vashon Glacier in the Puget Sound area receded (Campbell and Butler 2010; can assist in dating the associated assemblages through relative dating at a lower cost instead of McKechnie and Moss 2016). Salmonid species are incredibly instrumental to many indigenous tribes in radiocarbon dating. the Pacific Northwest; however, a variety of other fish, mammals, reptiles, amphibians, and shellfish are

The Washington State Department of Fish and Wildlife (WDFW) required any individual, company, important in ethnobiological records. One of the earliest archaeological analyses recorded in the or institution to apply for a Scientific Collections Permit (SPC) which the recipient must declare the Columbian Basin quantified species of animals from 8 sites along the Columbia River. The assemblages species, quantity, and intent of faunal collection. Antiquity Consulting chose to include animals on the included mammals such as moose (Alces alces), white-tailed deer (Odocoileus virginianus), mule deer permit that were introduced to western Washington on tracible dates such as the ring-necked pheasant (Odocoileus hemionus), elk, bison (Bison bison), mountain sheep (Ovis canadensis), mountain goat, black bear introduced in 1881, eastern gray squirrel introduced in 1925, and the nutria introduced in the 1930s (Ursus americanus), coyote, cougar (Puma concolor), bobcat (Lynx rufus), Canada lynx (Lynx canadensis), (Table 1). Other animals such as salmonid were included on the SCP to accurately identify certain fisher (Pekania pennanti), badger (Taxidea taxus), marmot, beaver (Castor canadensis), whistling swan (Cygnus species and taphonomic elements that could influence our understanding of the materials. columbianus), pileated woodpecker, dog, and whale (Collier 1942). Upon further study, faunal remains

#### **METHODS**

At the start of the project, Antiquity Consulting obtained information regarding methods of and potassium (Ferreira et al 2020). acquiring, skeletonizing, and preserving animals to complete the SCP application requirements. After consulting the Burke Museum, multiple associate professors of archaeology at multiple universities, dermestid beetles, natural burial and excavation, and water maceration. Dermestid beetle colonies and WDFW, and Washington State Department of Natural Resources, Antiquity Consulting prepared a study plan and submitted the application with a list of intended species to collect.

of each animal. Antiquity Consulting chose to use only one salvaged animal of each species that was until all organic material fell away from the bones (Hussain et al 2007; Sullivan and Romney 1999). When deceased without the company's intervention. Methods of collection include fish hatcheries at the end of changing the water, it was run through ½ inch, ¼ inch, and 1/8 inch mesh screens. The bones were spawning season, roadkill, chance encounter, hunters looking for carcass disposal options, pest control, placed in a diluted hydrogen peroxide solution for 3 to 10 days. Once in was completely dry, the and no-kill animal shelters (Post 2003). Antiquity Consulting recognized that although animals have and completed skeleton was catalogued and stored in an acid-free container for future comparative analysis will not be directly harmed due to the comparative collection, taking wildlife from an area before the (Elbroch 2006). carcass decomposed can deprived an ecosystem of reintroduced phosphorous, nitrogen, carbon, calcium,

Multiple methods of skeletal preservation were considered by Antiquity Consulting including burial both required facilities not available (Elbroch 2006; Hinshaw 2006), which left cold-water maceration as the best option within the company's capabilities. Once an animal was obtained, it was The application also required a statement concerning the method of collection and final disposition disarticulated and placed in a solution of water and soap that was changed with a screen every few days



Figure 1-3: Chinook salmon was collected from the Tumwater Falls Hatchery with permit. The first stage of processing the fish was gutting and separating the bones from the flesh (1). The bones are then placed in a combination of water and dish soap for 3 to 6 weeks. The water is periodically changed by screening the water to catch all the bones (2). After the process is complete, the skeleton was rearticulated to dry and store (3).





Figure 4-5: Townsend mole (Scapanus townsendii) before maceration process; anterior view (4) and posterior view (5).

#### RESULTS

obtained from the Tumwater Falls Hatchery and is in the process of cleaning a Townsend mole observed. In addition to the factors, salmon fins and other bonelike structures found in fillets are (Figure 4 and 5) in the 2024 SCP cycle. Of the two animals collected, there are various calcified nerve endings and cartilage, not bone. During the butchering process, many of these taphonomic marks such as carnivore gnawing present in the comparative collection providing components were most likely discarded without detection. Multiple elements of the skeleton valuable comparison of archaeologically associated faunal remains.

articulating the fish, components of the skeleton that had been lost during the maceration process maceration process. (Figure 3). The missing parts may have been the result of soaking the fish too long, small bones

Antiquity Consulting collected, preserved, cataloged, and stored a chinook salmon (Figure 1-3) slipping through all three sizes of screens, or the fish having decomposed more than initially required rehydration to prevent damage while rearticulating the animal.

Even with a completed chinook salmon skeleton, a few bumps in the road were encountered The Townsend mole is in the beginning stages of cold-water maceration. Due to the size of while it was processed. The salmon was much younger and more fragile than anticipated. While re- the rodent, a smaller container is used, and more time was taken to prepare the animal for the

### **DISCUSSION**

The company will create taphonomic markers such as charred bones and butcher marks to time and resources for a project. Companies may not have the time or resources to give better compare fauna from the archaeological record. Due to limited facilities, the focus of attention to faunal identification and taphonomy research when a comparative collection is the 2024 SCP cycle will focus on smaller animals such as fish, rodents, and small mammals. too far away or difficult to access. CRM has the responsibility to create resources to analyze Cold-water maceration, especially during the winter season, requires a longer period for the archaeological material to the best of our ability. Companies can create their own regional maceration process and more equipment when multiple animals are processed which should comparative collection through the permitting process along with a few rudimentary tools. be kept in mind before collection. An ideal fauna collection would have skeletons that were The faunal comparative collection that Antiquity Consulting is building will be available buried and underwent the natural process of decomposition to recreate a more accurate to local CRM companies to assist with faunal analysis. The company is also actively looking image of the taphonomy encountered in the archaeological record.

Cultural Resource Management (CRM) is a field which must balance protecting a complete animal listed on the poster.

Antiquity Consulting will continue to add animals listed in the permit to the collection. archaeological sites through employing careful methods of survey and analysis with limited

for sources of animals listed in Table 1. Contact Antiquity Consulting if there are remains of

| Table 1: 2024 Species Collection |                          |             |                      |                        |           |
|----------------------------------|--------------------------|-------------|----------------------|------------------------|-----------|
| SPECIES                          | SCIENTIFIC NAME          | COLLECTED   | SPECIES              | SCIENTIFIC NAME        | COLLECTED |
| chinook salmon                   | Oncorhynchus tshawytscha | $\boxtimes$ | great gray owl       | Strix nebulosa         |           |
| coho salmon                      | Oncorhynchus kisutch     |             | American robin       | Turdus migratorius     |           |
| chum salmon                      | Oncorhynchus keta        |             | house sparrow        | Passer domesticus      |           |
| house mouse                      | Mus musculus             |             | red-winged blackbird | Agelaius phoeniceus    |           |
| common opossum                   | Didelphis virginiana     |             | American crow        | Corvus brachyrhynchos  |           |
| eastern cottontail               | Sylvilagus floridanus    |             | California quail     | Callipepla californica |           |
| pika                             | Ochotona princeps        |             | black rat            | rattus rattus          |           |
| raccoon                          | Procyon lotor            |             | nutria               | Myocastor coypus       |           |
| striped skunk                    | Mephitis mephitis        |             | feral pig            | Sus sciofa             |           |
| western gray squirrel            | Sciurus griseus          |             | chicken              | Gallus gallus          |           |
| hoary marmot                     | Marmota caligata         |             | domesticated dog     | Canus familiaris       |           |
| black-tailed deer                | Odocileus hemionus       |             | house cat            | Felis catus            |           |
| elk                              | Cervus elaphus           |             | mountain goat        | Oreamus americanus     |           |
| coyote                           | Canis latrans            |             | domestic cow         | Bos taurus             |           |
| Townsend's mole                  | Scapanus townsendii      | $\boxtimes$ | horse                | Equus caballus         |           |
| Townsend's chipmunk              | Tamias townsendii        |             | ring-necked pheasant | Phasianus colchicus    |           |
| little brown bat                 | Myotis lucifugus         |             | pileated woodpecker  | Dryocopus pileatus     |           |
| long-toed salamander             | Ambystoma macrodactylum  |             | peregrine falcon     | Falco peregrinus       |           |